

WHAT MAKES RNA GENOMES SPECIAL?

SEARCH FOR THE HYDROGEN ATOM OF VIRUSES

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Honoring David Chandler, John Weeks, and Julia Yeomans

(SIMPLEST) VIRUSES ARE JUST:

A COMPOSITE OF

a nucleic acid genome (RNA or DNA)

AND

a protein shell -- “capsid”

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graph TD; A["a protein shell -- 'capsid'"] --> B[cylindrical]; A --> C[spherical]; B --> D[helical symmetry]; C --> E[icosahedral symmetry]
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cylindrical

helical symmetry

spherical

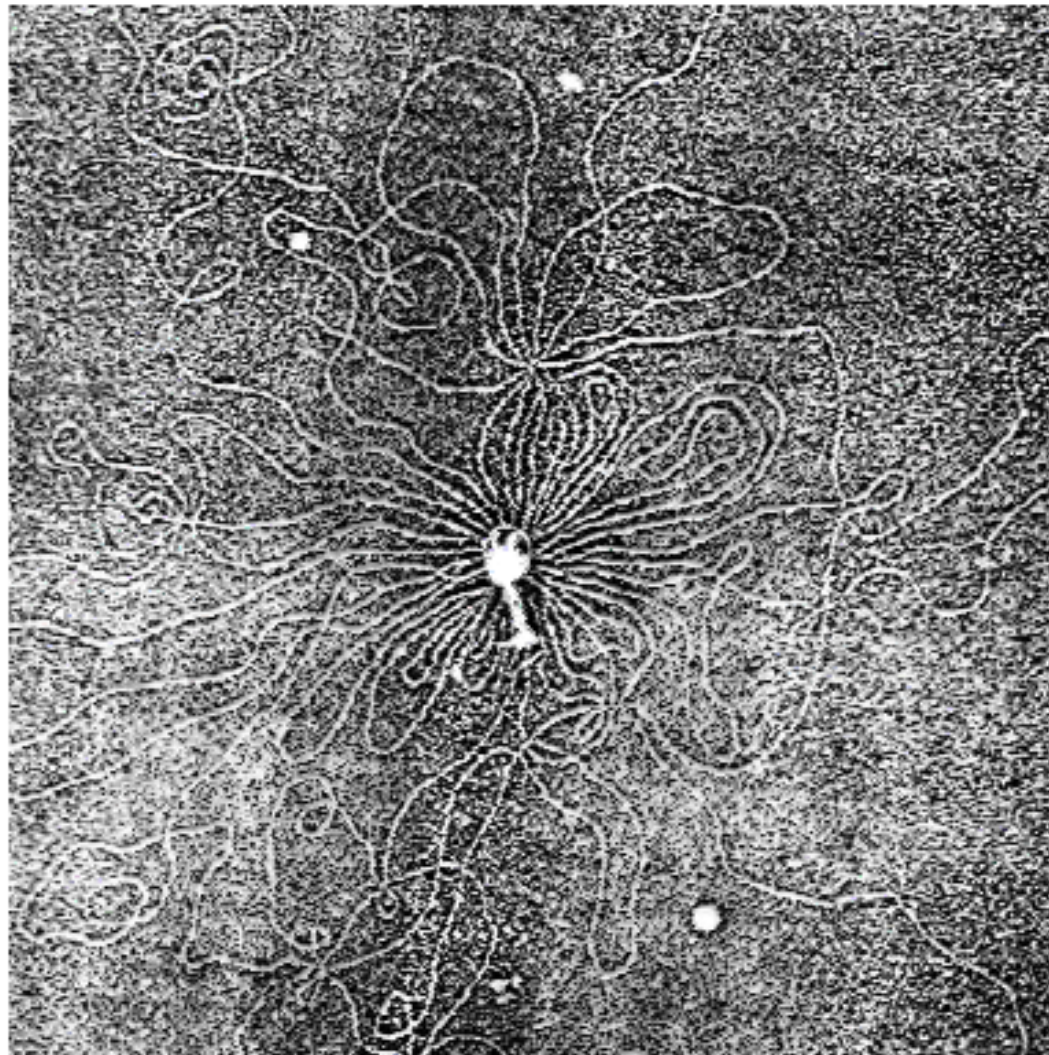
icosahedral symmetry

HOW ARE **RNA** VIRUSES DIFFERENT (FROM **DNA** VIRUSES)?

(single-stranded [ss]) **RNA** viruses (mostly plant and animal)
ssRNA is weakly confined; packaged spontaneously

(double-stranded [ds]) **DNA** viruses (mostly bacterial)
dsDNA is strongly confined; packaged by force

A **gene of DNA** is a very different *physical object* than a **gene of RNA**



Kleinschmidt et al. (1962)

50 nm

Osmotically-shocked
bacteriophage T2

$$R_{capsid} \approx 25nm \ll R_{DNA} \approx 1\mu$$

DNA “contour length” L , $\approx 20\mu$

DNA “persistence length” ξ , $\approx 50nm$

DNA “size” $(L\xi)^{1/2}$, $\approx 1\mu$

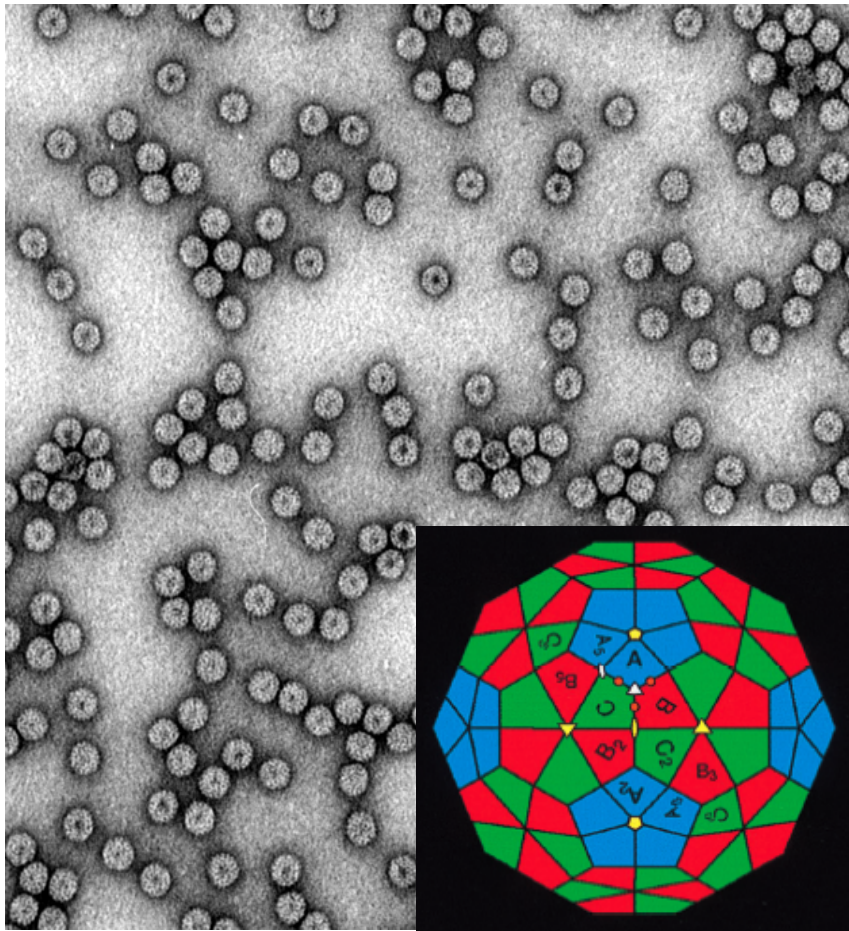
$$L \gg \xi$$

$$R_{DNA} \approx (L\xi)^{1/2} \sim M^{1/2}$$

LARGE DNAs ARE LINEAR
STATISTICAL OBJECTS WITH
WELL-KNOWN CHARACTERISTICS

**A LOT OF WORK HAS TO BE DONE, TO PACKAGE THE DNA GENOME
INTO A PRE-FORMED CAPSID – IT IS ... PRESSURIZED!**

What about viruses with *single-stranded (ss) RNA* genomes?



E.g., Cowpea Chlorotic Mottle Virus (CCMV)

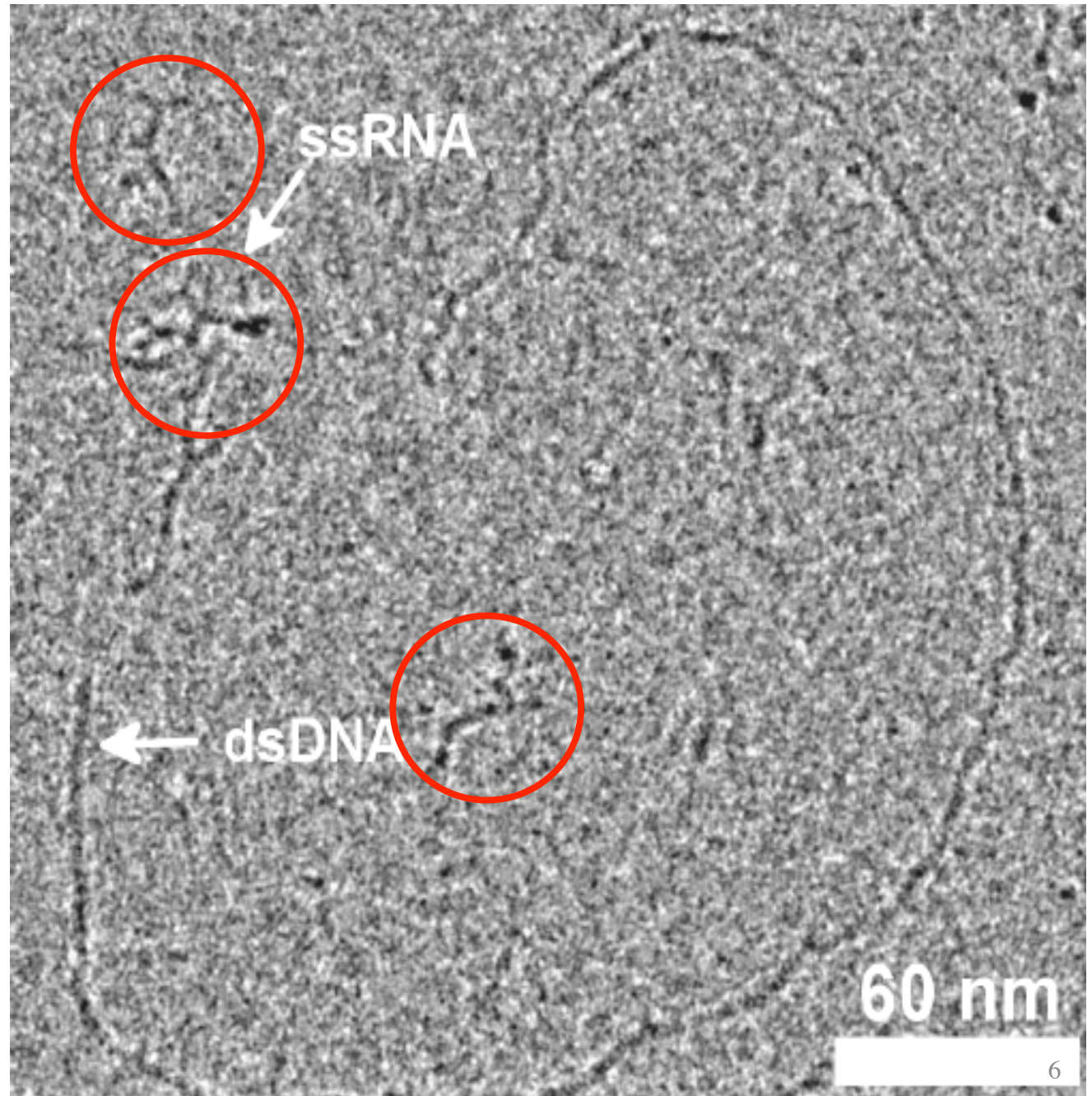
Each identical 28nm-capsid consists of *exactly* 180 copies of one protein, and contains a *different* molecule of the viral RNA genome – RNA1, RNA2, or RNA3 (+RNA4) – *each about 3000nt long*

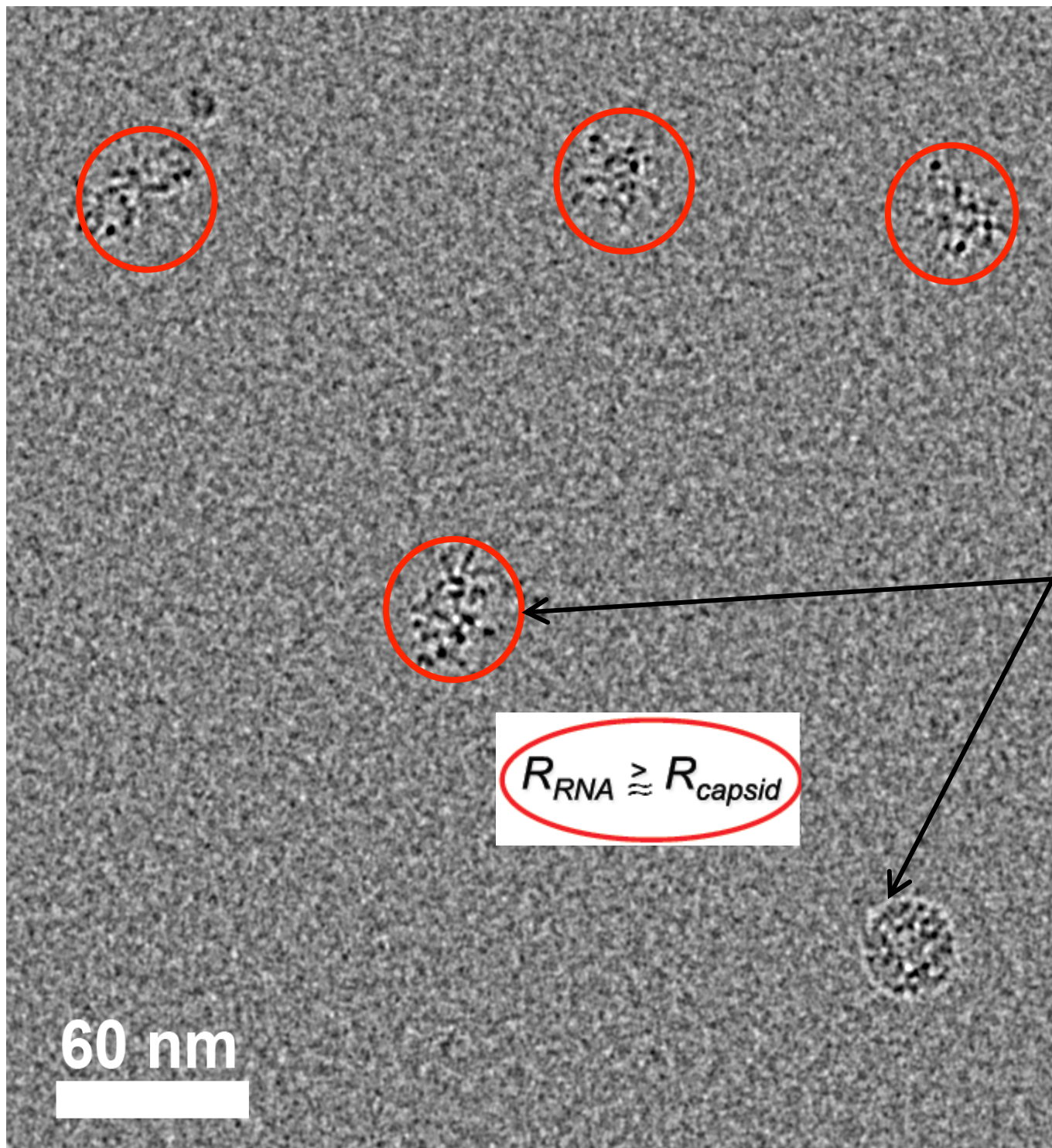
Packaging of genome occurs spontaneously,
via self-assembly – no work, no pressure – **WHY? HOW?**
BECAUSE ssRNA MOLECULES – GENES –
ARE TOTALLY DIFFERENT FROM THEIR DNA COUNTERPARTS...

**COMPARISON BETWEEN
2117 nt ssRNA
AND
2117 bp dsDNA**

Gopal, Zhou, Knobler, and
Gelbart
RNA **18**, 284 ([2012](#))

TE buffer pH 7.4





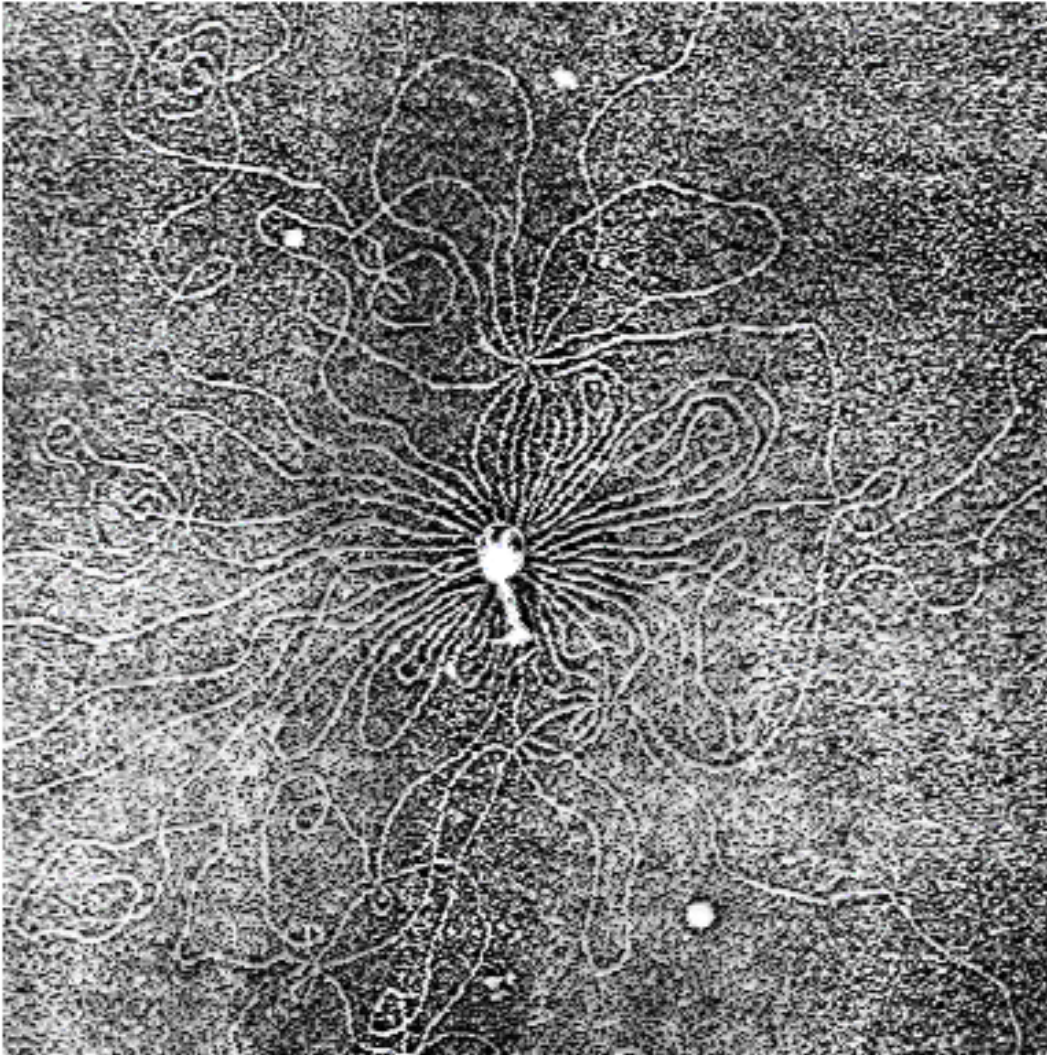
(cryo-EM micrograph)

**virion of CCMV, and
its gene content –
ss RNA2 (2774 nt)**

in assembly buffer
(physiological pH/ionic
strength, with Mg^{2+})

Gopal, Zhou, Knobler,
Gelbart, *RNA* (2012)

In contrast...



Large DNA is a stiff and linear, statistical object, taking up a lot of space, i.e., it is highly ramified

We know all its configurational properties, independent of sequence, if we know its contour length L and its stiffness ξ
Further,....

$$R_{DNA} \approx (L\xi)^{1/2} \gg R_{capsid}$$

NOT SO.....
for long RNA molecules

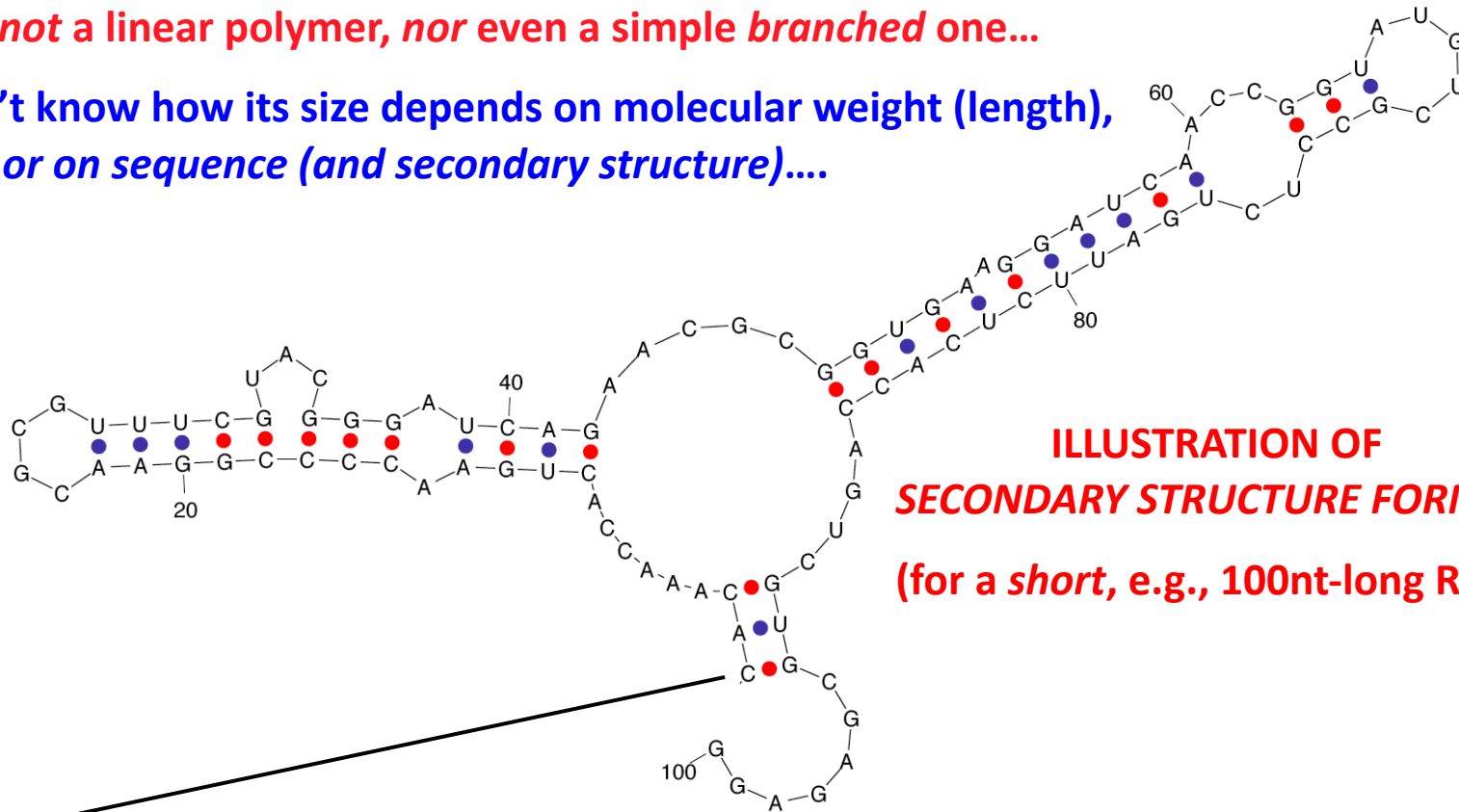
Kleinschmidt et al. (1962) Osmotically-shocked phage T2

Compactness of ssRNA accounts for smaller size of virus

What determines the size, shape, and flexibility of ssRNA?

It is *not* a linear polymer, *nor* even a simple *branched* one...

Don't know how its size depends on molecular weight (length),
or on sequence (and secondary structure)....



(5')CACAAACCACUGAACCCCGGAACGCGUUUCGUACGGGAUCAGAACGCGGUGAAGGA
UCAACCGGUAUGUCGCCUCUGAUUCUCACCAGUCGUGCGAGAGG(3')

But what about many *1000s-of-nt-long* RNA?

What *coarse-grained* features of its branching and shape and overall size
can we infer from its sequence?

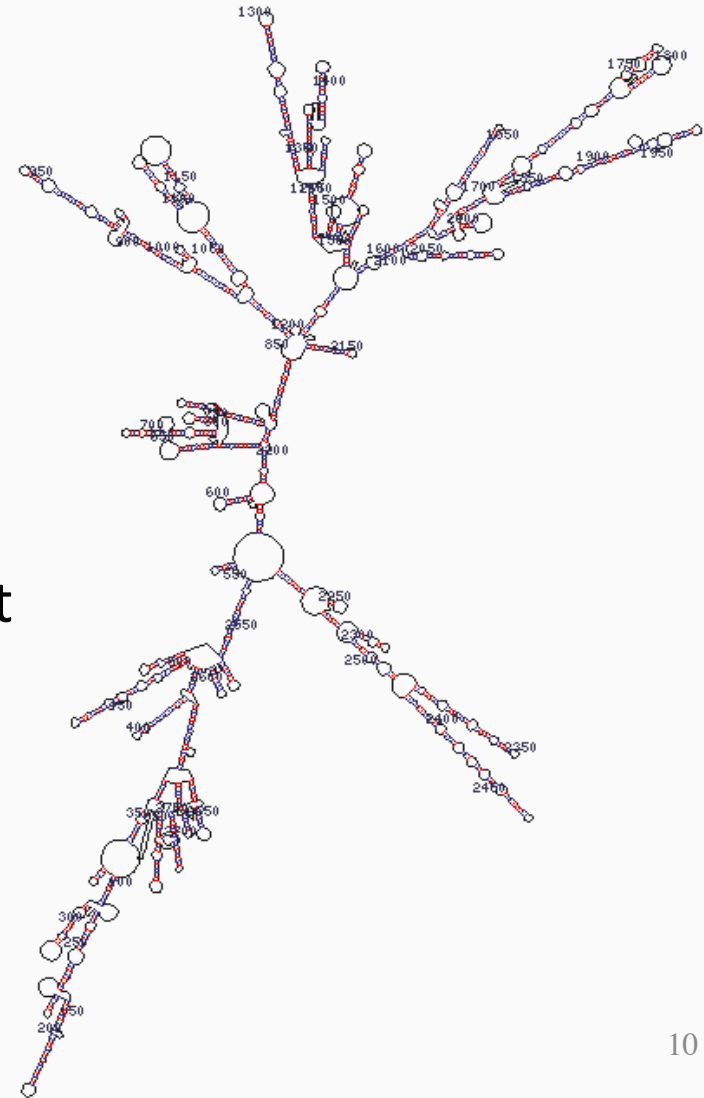
How does the *3D size and shape* of an arbitrarily long RNA correspond to its *secondary structure*?

1st example:

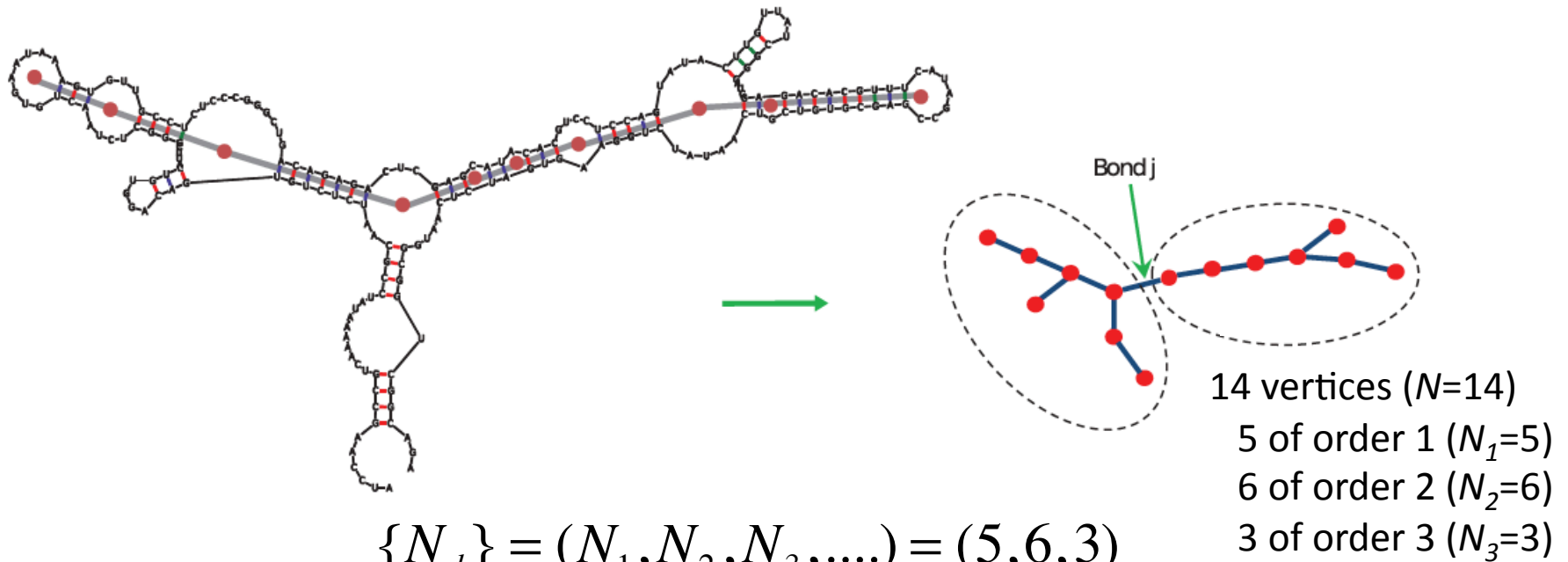
RNA2, the molecule comprising the second gene of CCMV, 2774 nt long

Is anything special about its secondary structure?

Can we identify some *coarse-grained* feature of its secondary structure that determines its overall 3D size?



Each RNA secondary structure can be mapped onto a “tree graph”
(and a *primary sequence* is mapped onto an *ensemble* of tree graphs...)



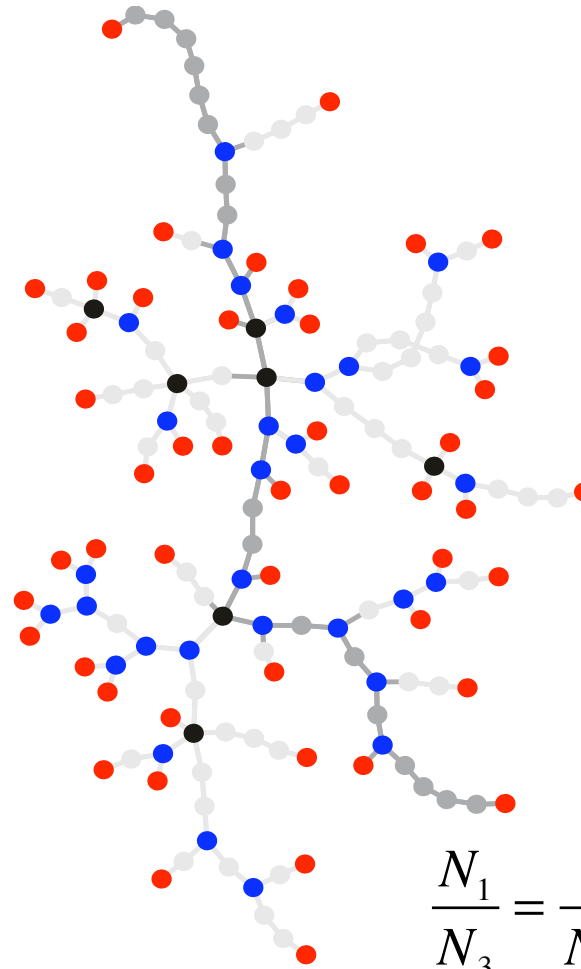
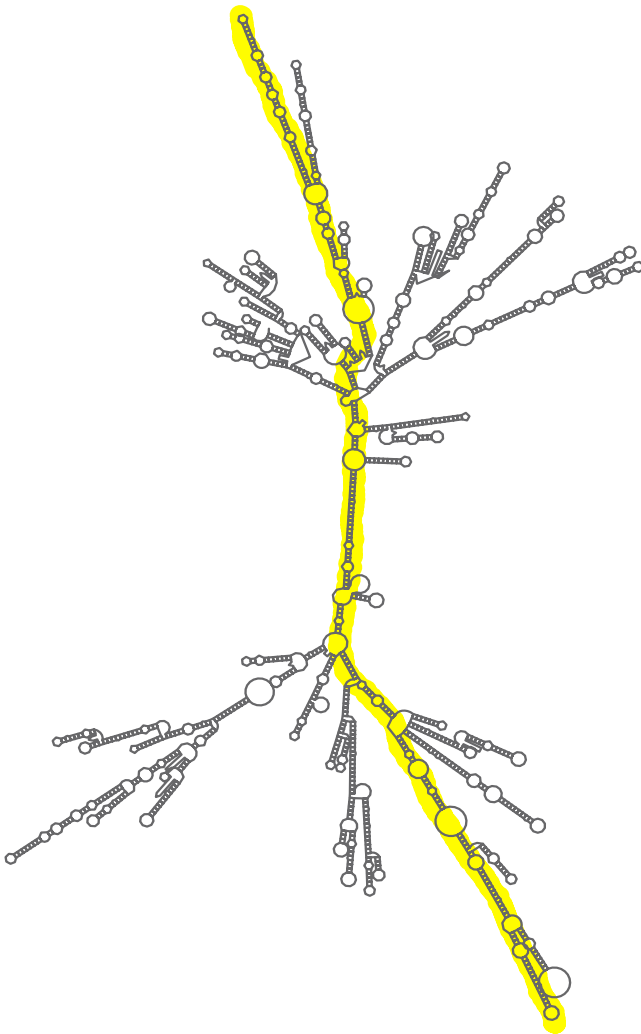
$$\{N_d\} = (N_1, N_2, N_3, \dots) = (5, 6, 3)$$

$$N = \sum_{d=1,2,3,\dots} N_d = 14$$

$$\langle R_g^2 \rangle = \frac{b^2}{N^2} \sum_{j=1}^{N-1} N_{\text{left}}(j) N_{\text{right}}(j), \quad b = \text{edge (Kuhn) length}$$

KRAMERS, 1940

Back to 2774 nt RNA2 of CCMV:



RNA Length		2774nt
%GC		42
Avg. Duplex Length including single-base bubbles.		5
T R E E	Total Vertices in Tree Graph	N=149
	Terminal Vertices (i.e. stem loops)	$N_1=48$
G R A P H S	2-fold Junctions	$N_2=64$
	3-fold Junctions	$N_3=30$
	4-fold Junctions	$N_4= 5$
	5-fold Junctions	$N_5= 2$
	6-fold Junctions	$N_6= 0$ *

$$\frac{N_1}{N_3} = \frac{2}{N_3} + 1 + 2 \frac{N_4}{N_3} + 3 \frac{N_5}{N_3} + \dots$$

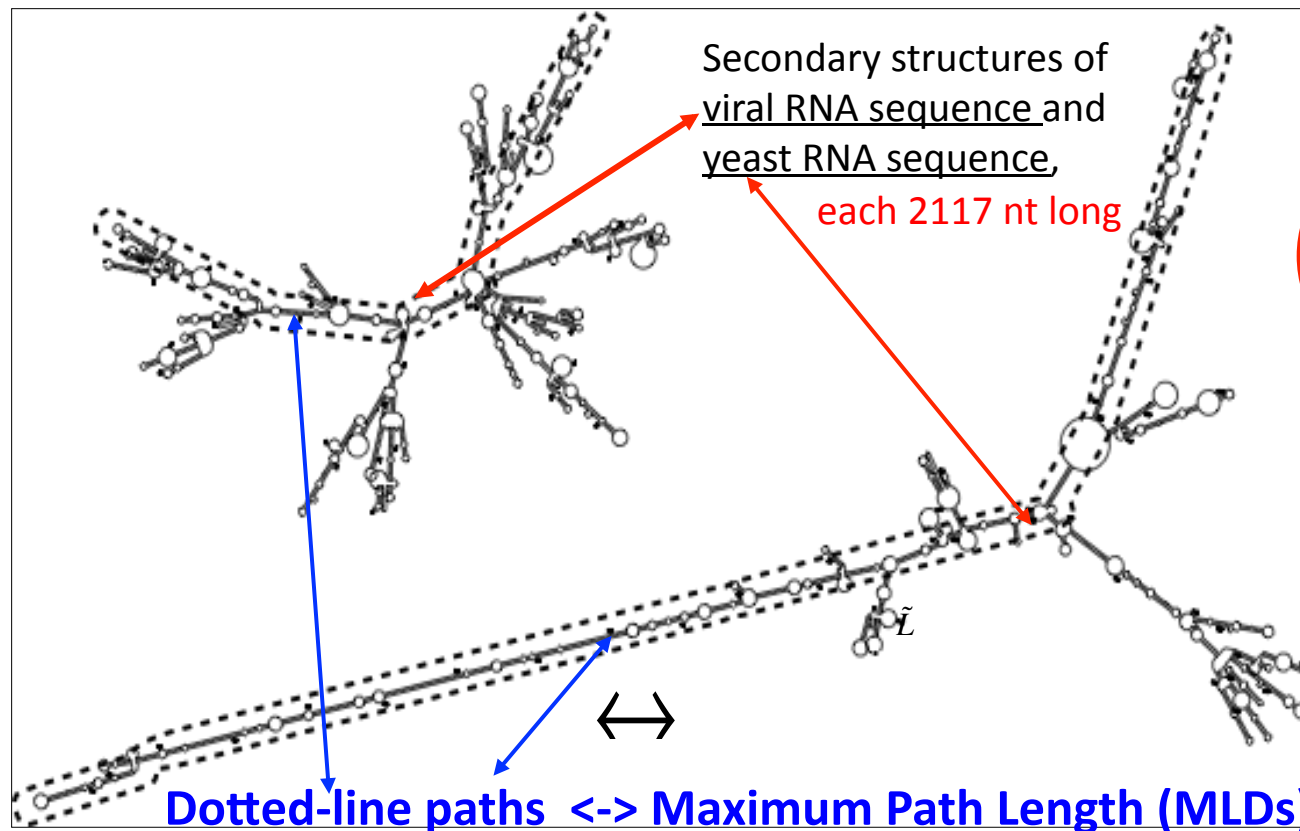
EULER, 1750

LONG RNA IS A FLEXIBLE, BRANCHED, STATISTICAL OBJECT WITH UNKNOWN CHARACTERISTICS

(E.g., we don't know the significance of the *distribution of vertex orders*, $\{N_d\}$)

For long RNAs: $N_3 \gg 1 \Rightarrow \left(\frac{N_1}{N_3} - 1\right)$ is a measure of *higher-order branching ... compactness*

Another example: RNA 3, the third molecule (2117 nt) of the CCMV genome



Viral sequences have smaller MLDs, implying smaller R_g s, and hence are *more compact*

Yoffe, Prinsen, Gopal, Knobler, Gelbart, Ben-Shaul, *PNAS (USA)* **105**, 16153 (2008)

WHAT ACCOUNTS FOR THE DIFFERENCES IN THEIR 3D SIZES?

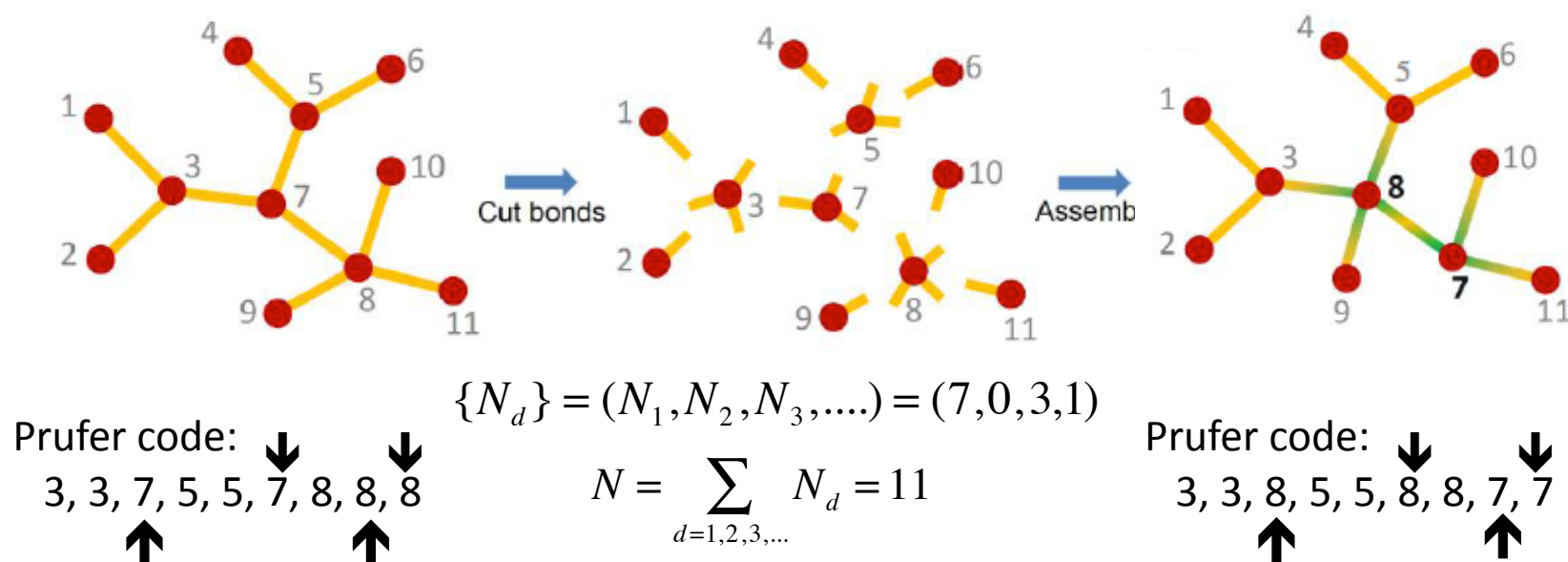
E.g., treat RNA as a *linear homopolymer* with $\tilde{L} \leftrightarrow MLD$:

$$R_g \approx (\tilde{L} \tilde{\xi})^{1/2} \rightarrow (MLD \tilde{\xi})^{1/2} \propto MLD^{1/2}$$

Here \tilde{L} is the (*effective*) contour length, and $\tilde{\xi}$ is the (*effective*) persistence length

**BOTH STRUCTURES HAVE COMPARABLE BRANCHING, E.G., SIMILAR $\{N_d\}$ s,
BUT ONE HAS A MORE COMPACT CLUSTERING OF BRANCHING POINTS...**

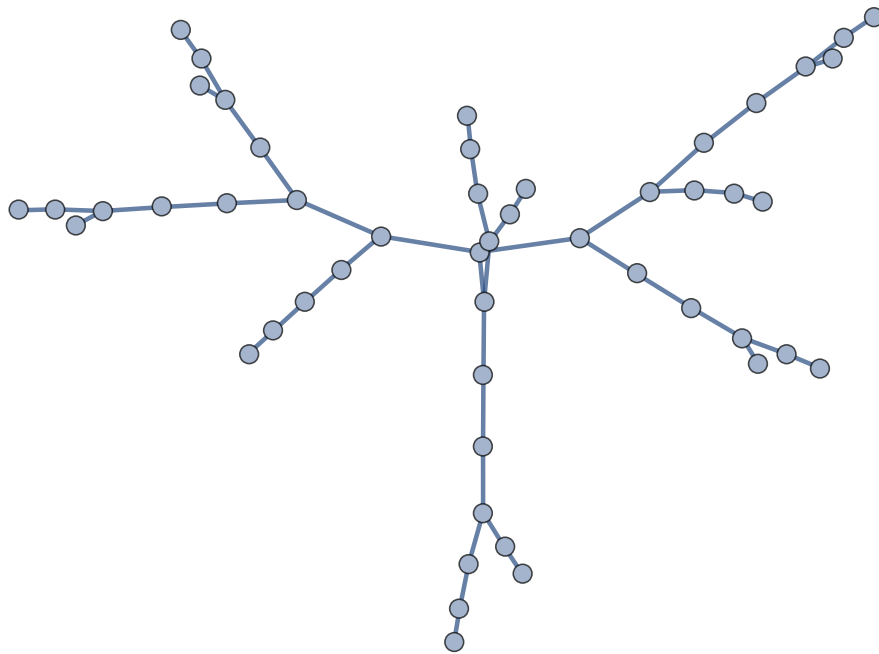
PRÜFER, 1918 : AN ARBITRARY TREE GRAPH CAN BE REPRESENTED BY
A UNIQUE ORDERED SEQUENCE OF INTEGERS – “THE PRUFER CODE”



Here we have “shuffled” the Prüfer sequence by permuting two pairs of integers

Prüfer shuffling: Leaves invariant the number of vertices (N), and the distribution of vertex orders $\{N_d\}$...but changes the connectivity of the graph, and hence its “size” (compactness)

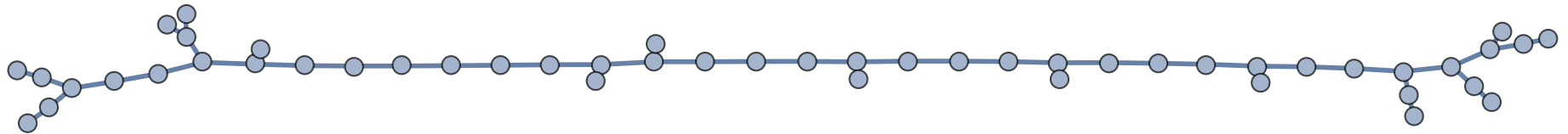
Singaram, Garmann, Knobler, Gelbart, Ben-Shaul, *J. Phys. Chem. B* **119**, 13991 (2015)



These two tree graphs have the
*same number of vertices, and the
 same distribution of vertex orders,*

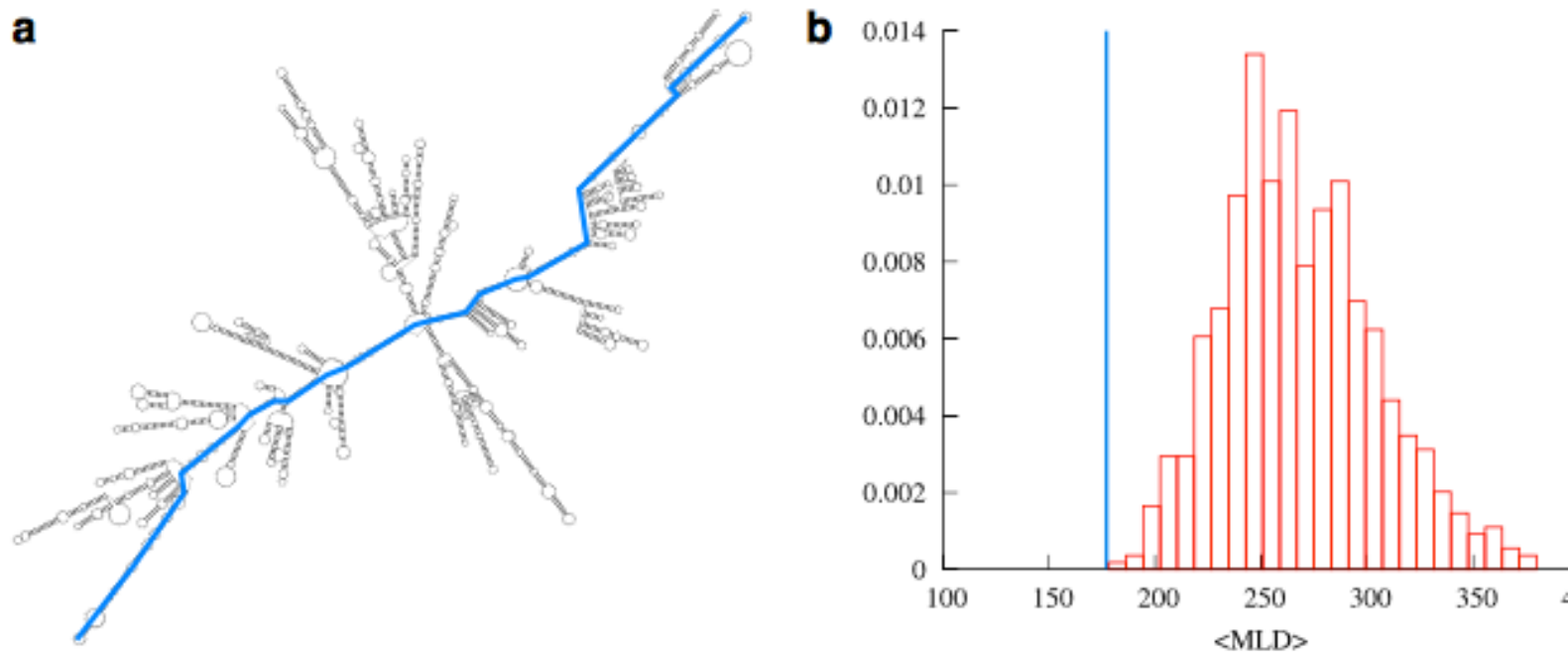
$$\{N_d\} = (N_1, N_2, N_3, \dots) = (14, 24, 12)$$

$$N = \sum_{d=1,2,3,\dots} N_d = 50$$



The 1st has been obtained from the 2nd by successive permutations of its Prufer sequence,
*chosen to decrease the maximum path length (MLD),
 and hence its radius of gyration, or – equivalently – the compactness of its branch points*

VIRAL SEQUENCES ARE MORE COMPACT...



a Typical secondary structure of BMV RNA2, with the maximum path length (MLD) shown in **blue**.

b Thermally-averaged maximum path length (**blue line**) of the viral (RNA2) sequence, and the distribution (see **red histogram**) of averaged maximum path lengths for each of many random sequences with the same length and nt composition.

VIRAL RNA GENOMES HAVE EVOLVED – NOT JUST TO CODE FOR CERTAIN PROTEINS – BUT ALSO TO BE MORE COMPACT

Viral sequences are more compact....

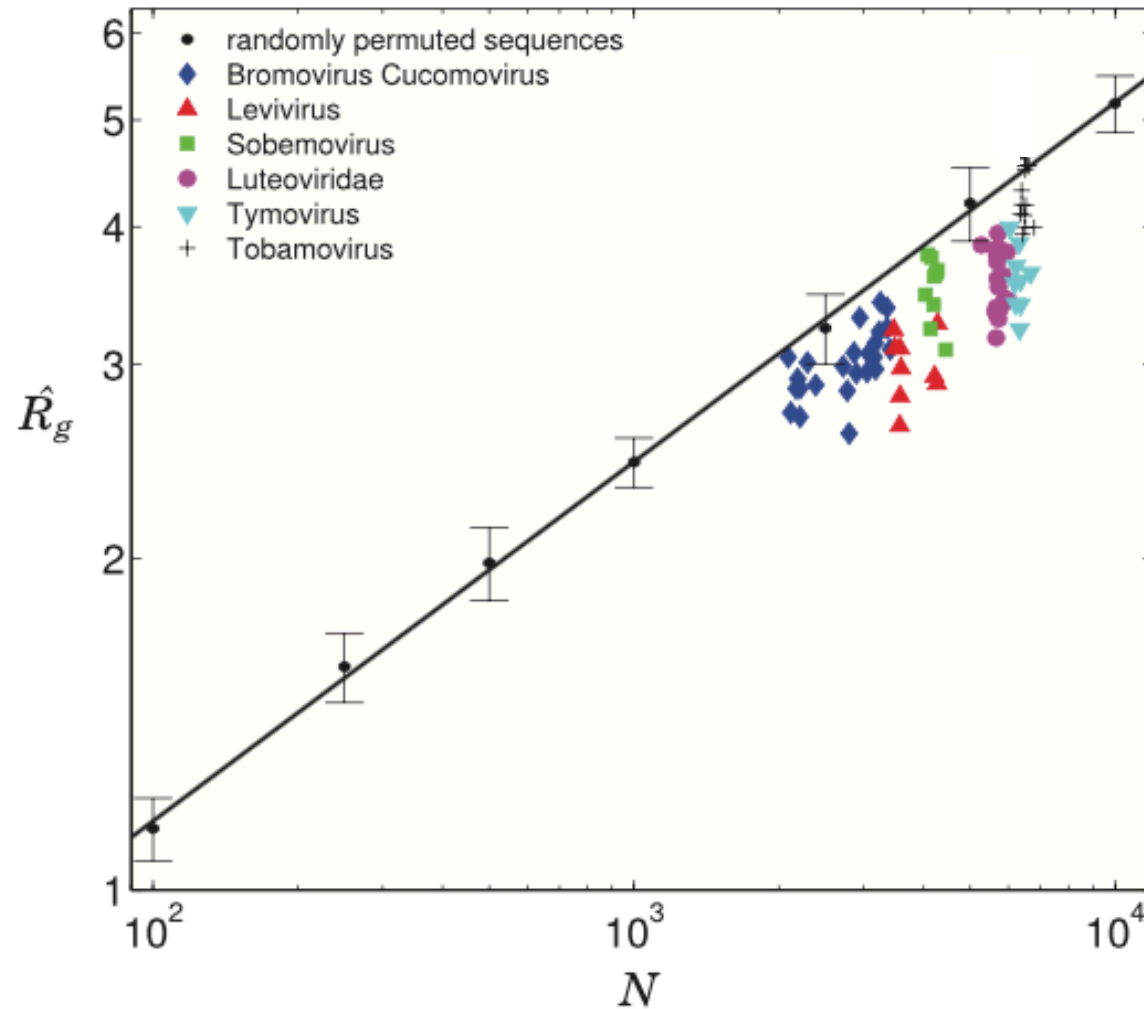
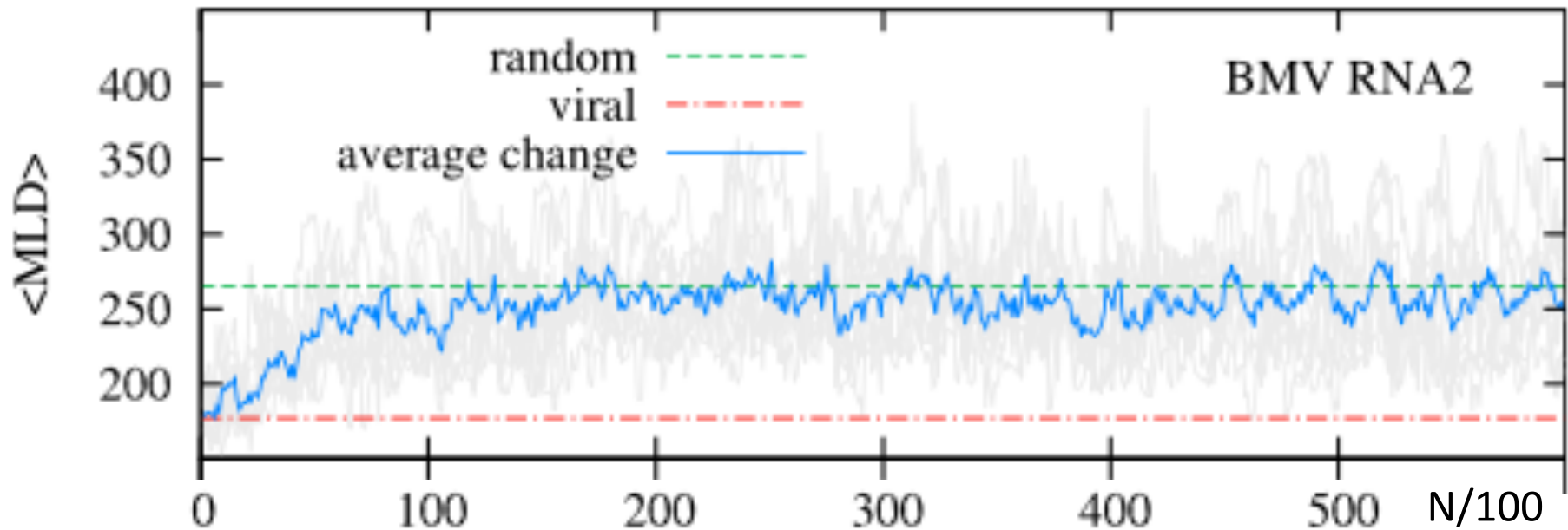


FIG. 3. A log-log plot of the radius of gyration, \hat{R}_g (in units of segment length, b), as a function of RNA sequence length, N . Each black dot represents the average result obtained for 20 randomly shuffled sequences of equal base composition.

SYNONYMOUS MUTATIONS HAVE BEEN RECRUITED TO COMPACTIFY VIRAL SEQUENCES



Average maximum path length (<MLD>) after N synonymous mutations have been introduced into the viral (CCMV RNA2) sequence, for 10 trajectories (grey) and their average (blue)

Red dot-dash line: average value for viral sequence

Green dashed line: average value for random sequences with same length and nt composition

Tubiana, Bosc, Micheletti, Podgornik, *Biophys. J.* **108**, 194 (2015)

Ben-Shaul, Gelbart, *Biophys. J.* **108**, 14 (2015)

RNA VIRUSES ARE SMALLER THAN DNA VIRUSES, because
(physical reason) RNA genomes are more compact, per gene
(biological reason) RNA viruses have fewer genes (smaller genomes)

- Compactness of RNA genomes derives from their being (effectively) highly branched polymers

And viral RNA is more *compactly* branched (*smaller!*) than non-viral RNA, thereby competing better for binding (and packaging) by capsid protein

- *How few genes can an RNA virus have?*

CAN WE MAKE A VIRUS WITH ONLY 2 GENES...?

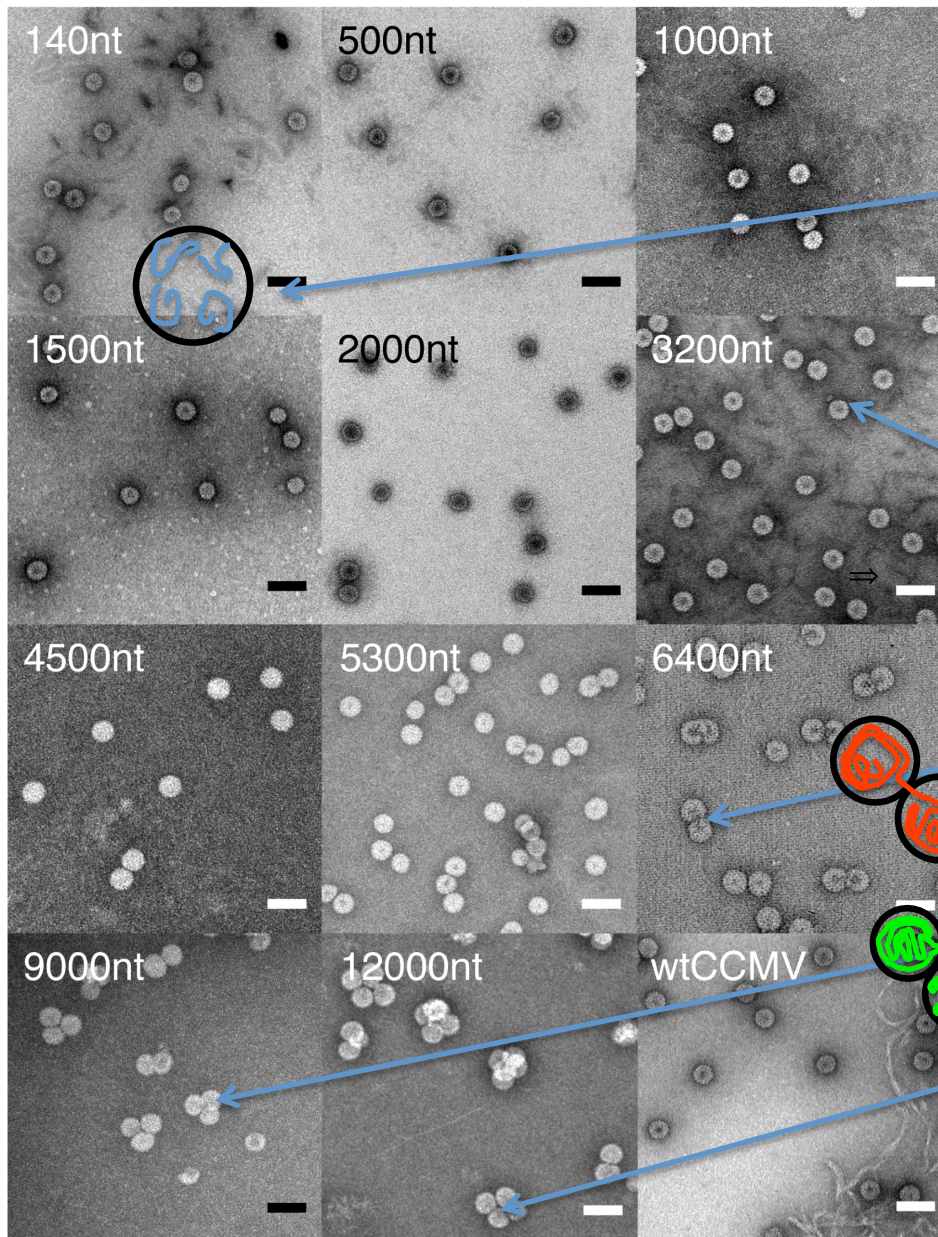
Work done with

Charles Knobler [UCLA] and Avinoam Ben-Shaul [Jerusalem]

Dr. Ajay Gopal

Walter Singaram

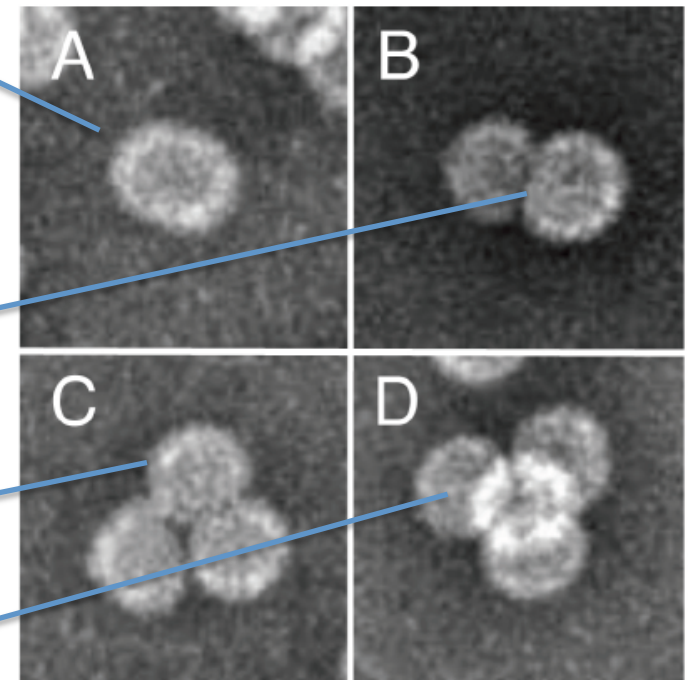
We (i.e., CCMV protein) can package – *in vitro* – any RNA, of any length:



“undersized” RNAs \Rightarrow
many RNAs per capsid

“oversized” RNAs \Rightarrow
many capsids per RNA

(about 3000 bases per capsid)



Cadena-Nava, Comas-Garcia, Garmann, Rao,
Knobler, Gelbart, *J. Virology* **86**, 3318 (2012)

Capsid protein subunits form closed, icosahedrally-symmetric, 2D hexagonal lattices:
Euler's 12 five-fold defects appear as icosahedrally-positioned pentamers

Minimum-energy capsid structures correspond to those with the *minimum number* ($T=1, 3, 4, 7, \dots$) of *inequivalent positions* for the $60T$ protein subunits

$T=1$



$T=3$



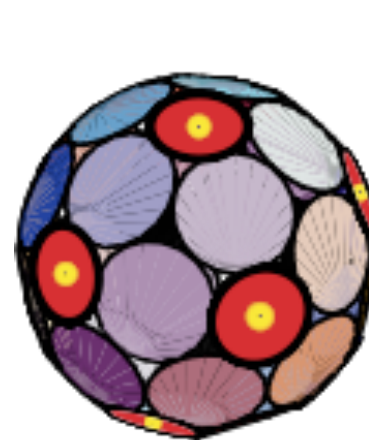
These structures self-assemble, spontaneously, around ssRNA...!

$N=12$: 12 pentamers,
 $0=10(T-1)$ hexamers
 $60T=60$ proteins, $T=1$

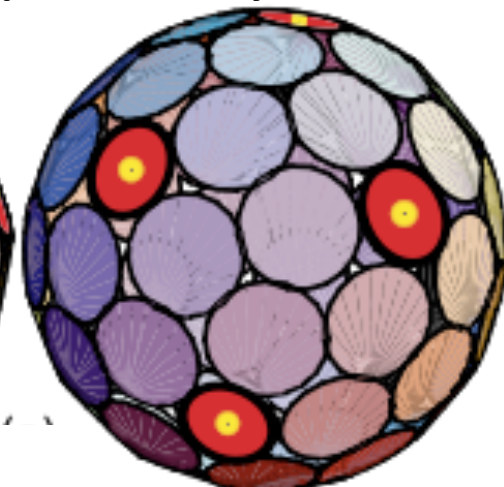
$N=32$: 12 pentamers,
 $20=10(T-1)$ hexamers,
 $60T=180$ proteins, $T=3$

$N=42$: 12 pentamers,
 $30=10(T-1)$ hexamers,
 $60T=240$ proteins, $T=4$

$N=72$: 12 pentamers,
 $60=10(T-1)$ hexamers,
 $60T=420$ proteins, $T=7$



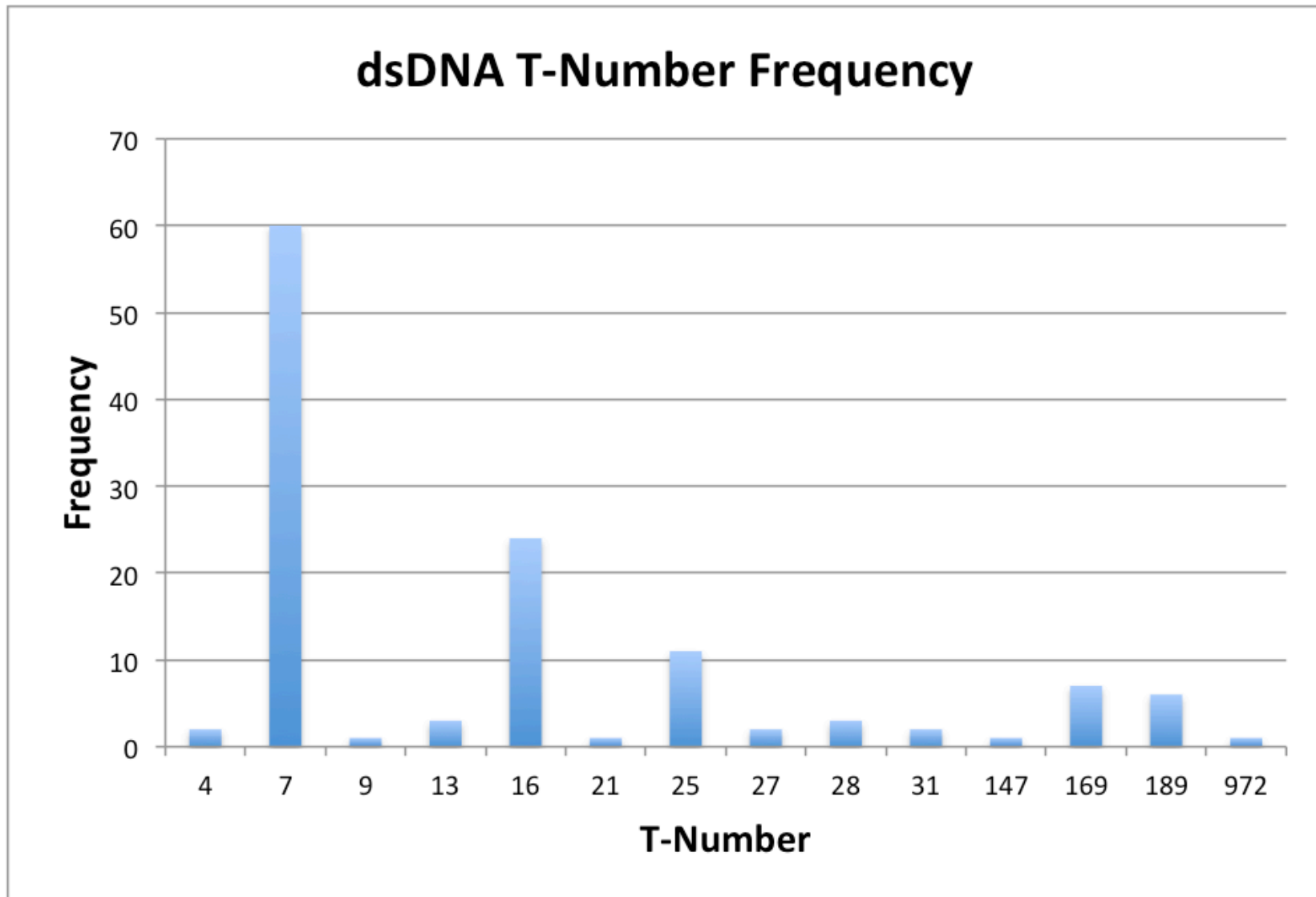
$T=4$



$T=7$

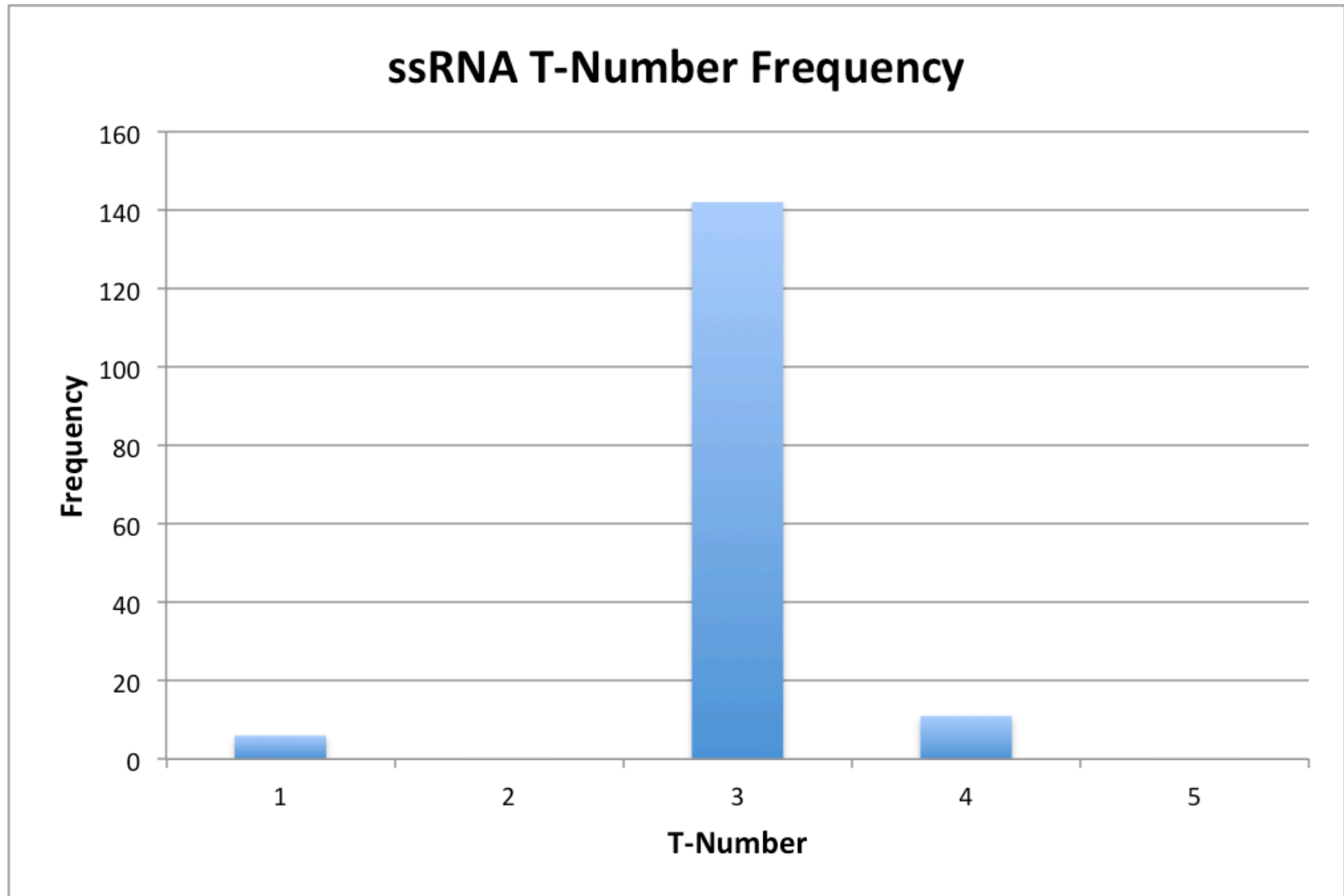
$$T = h^2 + k^2 + hk, \quad h, k = 0, 1, 2, \dots$$

Zandi, Reguera, Bruinsma, Gelbart, Rudnick, *PNAS (USA)* 101, 15556 (2004)



$$R_{capsid} \sim T^{1/2}$$

Liya Oster



(Typically, ssRNA viruses have 10-100 times fewer genes than dsDNA viruses!)